

<https://doi.org/10.33003/jaat.2023.0902.06>

NANOPARTICLES ANTIBIOTIC DELIVERY SYSTEM: AN ALTERNATIVE TREATMENT PROTOCOL FOR CASEOUS LYMPHADENITIS.

¹Idris Sherifat Banke and ²Arifah Abdul Kadir,

¹Department of Veterinary Pharmacology and Toxicology, Faculty of veterinary Medicine, Usmanu Danfodiyo University, Sokoto State, Nigeria.

²Department of Veterinary Preclinical Studies, Faculty of Veterinary Medicine, Universiti Putra Malaysia, 43400, UPM, Serdang, Selangor.

*Corresponding author Email : arifah@upm.edu.my Phone: +2348136103222

ABSTRACT

Caseous lymphadenitis is an infectious zoonotic disease of goats with severe economic implications. A major reason why management of the disease is a problem in the small ruminant livestock industry is because *Corynebacterium pseudotuberculosis* forms biofilm in the host and is resistant to antibiotic drugs resulting in a chronic granulomatous condition. The use of nanoparticles as a means of direct drug delivery to this organism may provide a solution to tackling it. This review focuses on caseous lymphadenitis and the different nanoparticles for antimicrobial drug delivery

Keywords: caseous lymphadenitis, antibiotic resistance, nanoparticle, drug delivery

INTRODUCTION

Caseous lymphadenitis caused by *Corynebacterium pseudotuberculosis* is a disease of severe zoonotic and economic importance in the small animal livestock industry (Galvao *et al.*, 2017). It is difficult to treat with antibiotics because *Corynebacterium pseudotuberculosis* evades the hosts immune system allowing it to multiply within the host tucked away in a pool of abscess surrounded by a thick fibrous capsule therefore making it difficult for the conventional formulations of antibacterial agents to get to the organism (Williamson, 2001). A number of approaches have been used in the management of CLA which includes administration of vaccine of the strain isolated from the herd, lancing and flushing with antibiotics, surgical removal of the abscess with identification and culling of affected animals however persistence of the disease in a herd even after long vaccination programs, recurrence of abscess following lancing and surgical removal and resistance to conventional antibiotics pose significant obstacles to achieving optimized therapeutics (Oreiby, 2015). The use of nanoparticles in the delivery of antibiotics to target sites is advantageous because it enhances pharmacokinetics and improves biodistribution of the loaded antibiotics whilst maximizing their delivery to target tissues, recently, Gold nanoparticles have been shown to demonstrate intracellular antibacterial activity against *C. pseudotuberculosis* (Mohammed *et al.*, 2017)

The delivery of antimicrobials using nanoparticles is better compared to the conventional methods of antimicrobial delivery since the bacterial outer (membrane plays a role in permeability of these drugs, hence improving antibacterial activity (Torres *et al.*, 2012). This review will dwell on caseous

lymphadenitis and nanoparticles for antimicrobial delivery.

METHODS

The databases used to search for articles used in this review includes peer reviewed journals on nanoparticle drug delivery published in PubMed and Medline. References from these articles were also examined for additional articles.

Caseous lymphadenitis

Caseous lymphadenitis (CLA) is a chronic suppurative disease of cattle, sheep and goats, characterized by palpable abscess or granulomatous nodules containing cheesy exudates in lymph nodes, skin and/or internal organs (Oreiby, 2015). CLA manifests with external lesions or visceral lesions. These two forms may occur together or separately in the same animal (Al-Gaabary *et al.*, 2009). In the external form of CLA, a distinct clinical manifestation is the swelling of external lymph nodes especially that of the parotid, submandibular, and supra mammary nodes though other lymph nodes may be involved. The internal form, it is characterized by chronic weight loss, coughing and other respiratory signs such as dyspnoea, tachypnea and nasal discharge. The clinical signs seen in either forms of CLA is usually related to which internal organ is affected (Habus *et al.*, 2015). Also, the disease may be asymptomatic with internal lesions only that may lead to chronic diseases related to the internal organs affected usually detected at post mortem (Ribeiro *et al.*, 2013).

Aetiology

Corynebacterium pseudotuberculosis is a Gram positive pleomorphic, microaerophilic (5% CO₂), intracellular bacterium which is neither encapsulated nor motile (Oreiby *et al.*, 2015; Nassar *et al.*, 2015).

The presence of two virulence factors facilitates its pathogenicity: phospholipase D (PLD), an exotoxin which enhances its spread by weakening endothelial cells and increasing vascular permeability and a mycolic acid rich toxic cell wall which protects it from host enzymes and prevents entry of antibiotics (Mahmood *et al.*, 2015). Two biotypes of the bacteria have been identified namely the nitrate reducing group which affects sheep and goats and the non-nitrate reducing group which affects horses. Cattle is affected by both groups (Washburn, 2018).

Pathogenesis

Corynebacterium pseudotuberculosis gains entry into the body mostly through the intra dermal route either by scratches, wounds, cuts, or via other less common routes like inhalational or intraperitoneal. Upon entry, the bacteria activate the immune system and phagocytes are attracted to that area. The phagocytes transport the bacteria to the lymphnodes through the regional draining lymphatic system (Bastos *et al.*, 2012). The bacteria replicates in the lymphnodes leading to the development of a pyogranuloma (as the external lipid coat protects it from the phagocytic action of the phagocytes.) The abscess becomes bigger and it is walled in a thick fibrous capsule which prevents further actions from the host immune system therefore making it inaccessible to antibiotics (Bastos *et al.*, 2012; Habus *et al.*, 2015; Washburn, 2018). The abscess formed is not limited to the lymph nodes alone, it can also be found on internal organs like liver, kidney, spleen and lungs.

Diagnosis

Diagnosis is based on presenting characteristic lesions of external abscess (Nassar *et al.*, 2015) and this may be challenging because some infected animals may be asymptomatic for a long time thereby facilitating the disease spread in the herd. CLA is usually confirmed by isolation, culture and biochemical identification of *C. pseudotuberculosis* to differentiate it from other bacterial organisms that are characterized by formation of pus in goats though this may not be efficient in chronic infection where the external lesions contain little pus and viable organisms (Ribeiro *et al.*, 2013). Serological test like ELISA and PCR are more efficient in diagnosis because they can be used to detect asymptomatic infection in a herd (Ribeiro *et al.*, 2013; Hariharan *et al.*, 2015; Nassar *et al.*, 2015)

Treatment

Treatment with antibiotics have been ineffective (Hariharan *et al.*, 2015) though other methods such as administration of vaccine of the strain isolated from the herd, lancing, flushing and surgical removal of abscess with identification and culling of affected animals have been used (Washburn *et al.*, 2013). Though these methods involve heavy economic implications (Washburn *et al.*, 2013). Treatment with antibiotics does not completely clear the infection and this may not be unrelated to the mycolic acid rich toxic cell wall and

bacterial virulence factors (Mahmood *et al.*, 2015; Washburn *et al.*, 2013). Experimental intralesional and systemic administration of a new macrolide antibiotic tulathromycin demonstrated positive results in goats when this antibiotic was used in flushing of lesions (Washburn *et al.*, 2013).

Resistance of *C. pseudotuberculosis* to antibiotics

Antibiotics are effective in treating most bacterial infections and cost effective unfortunately, unregulated, widespread use and misuse have led to the emergence of multidrug drug resistance strains. *Corynebacterium spp* have been demonstrated to develop resistance genes against most of the common antibiotics used for treatment (Oleander, 2012). *Corynebacterium pseudotuberculosis* forms biofilm which the animal's immune system cannot get rid of with resultant damages to surrounding tissues in trying to curtail its spread (Sa *et al.*, 2002). High microbial virulence and resistance to antibiotics are the major differences between planktonic and biofilm (sessile) forms of bacteria (Jamal *et al.*, 2018). Which makes biofilm associated infections difficult to treat as they do not yield to treatment with antibiotics which have been proven successful against the planktonic forms (Kostakioti *et al.*, 2014). The bacteria acquire resistance by: developing genes responsible for resistance, modification of the ribosome binding site associated with methylation or mutation, effective pumping active efflux of antibiotic out of the organism and enzymatic inactivation of the antibiotics (Oleander, 2012). Most antibiotics do not get access to the organism due to the development of bacterial biofilms which can evade the immune system, thus results in persistence and development of chronic infections (Wang *et al.*, 2017).

Nanoparticle antibacterial mechanism of action

Nanoparticle antibacterial mechanism of actions are not related to and does not depend upon any of those mentioned above as it involves direct contact with the bacterial cell wall, where it penetrate and destroy the bacterial generates free radicals which interact and interfere with cellular macromolecules and the direct reaction with functional groups of proteins, lipids and genetic materials (Wang *et al.*, 2017). Nanoparticles increases the rate of reactive oxygen species (ROS) formation in bacterial cells which is one way the nanoparticles exert their antimicrobial effects (Khan, 2017). The high surface area to volume ratio and small size characteristic of nanoparticles qualify them as suitable candidates in the treatment of infections caused by biofilms as this facilitates closer interactions with bacterial membranes. Nanoparticles are highly advantageous in the target drug delivery systems because they bypass bacterial resistance mechanisms to conventional antibiotic treatment thereby decreasing virulence and eradicating resistance as well as reducing cost expended on antibiotics in the treatment of infections (Ranghar *et al.*, 2014). Their ultra-small size

makes them have a higher surface to volume ratio with increased number of active atoms at their outer surfaces, makes contact with target site or organism better compared to large scale molecules (Mohamed *et al.*, 2017). It is engineered in such a manner that it has a more rapid onset of therapeutic action, non-immunogenic and are cost effective taking into consideration the effect of physicochemical properties such as pH, osmotic pressure and temperature (Aruguete *et al.*, 2017).

Types of nanoparticles for drug delivery

Inorganic nanoparticles

Inorganic nanoparticles are those nanoparticles that are free of carbon in their composition and they are usually metal or metal oxides based. Metal oxide based nanoparticles such as iron oxides (magnetite (Fe_2O_4) and maghemite (Fe_2O_3) (Chertok *et al.*, 2007), Aluminium oxide (Al_2O_3), Silicon dioxide (SiO_2) Zinc oxide (Mishra *et al.*, 2017) have been used as drug carriers for due to their biocompatibility and antimicrobial activity (Mukherjee *et al.*, 2011).

Metallic Nanoparticles

These are nanoparticles with inherent intrinsic optical, supra magnetic and biological properties. These optical and supramagnetic potentials have been harnessed in micro systems such that they can be used for highly technical tests in medical devices which is very rapid with high volume and minimal cost (Ladj *et al.*, 2013). Nanosized metals are within a size range of 1-100 nm. Their physicochemical properties and functionality have been optimised such that ligands such as sugars, peptide, protein and DNA can be linked to their surface (Bhatia, 2016). They have been used in biomedicine for the delivery of drugs and biosensors. Metallic nanoparticles like superparamagnetic iron oxide nanoparticles, zinc oxide nanoparticles, copper oxide nanoparticles, gold nanoparticles, titanium dioxide nanoparticles, gallium based nanoparticles which have been reported to have antibacterial activity, antiviral and antiparasitic activities (Aderibigbe *et al.*, 2017; Mohamed *et al.*, 2017). Gold and titanium dioxide nanoparticles also have been reported as resonance imaging contrast enhancers and anticancer agents carriers (Khanna *et al.*, 2015)

Quantum dots

These are nanocrystals of few nanometres consisting of pure heavy metals or a mixture of metals. It is commonly composed of selenium or cadmium or a mixture of both. They are used both for *in vitro* biomedical labelling in complex media due to their photostability and fluorescent properties (Ladj *et al.*, 2013).

Organic nanoparticles

Polymeric nanoparticles

These are solid colloidal nanoparticles made of biodegradable and biocompatible polymers and lipids (Mallakpour and Behranvand, 2016). They are in form of nanospheres or nanocapsules depending on the method of preparation. In nanocapsules, the drug is contained within the cavity surrounded by the polymer whereas in the nanosphere, the drug and the polymer are embedded in a drug polymer matrix (Muhamad and Selvakumaran 2014). Generally two polymers are used in preparation of polymeric nanoparticles, the naturally hydrophilic polymers (Proteins and polysaccharides) and the synthetic hydrophobic polymers which can either be pre-polymerized or polymerized. Natural polymers such as cellulose, starch, chitosan, carrageenan, alginates, xanthan gum, gellan gum and pectins have been used as drug carriers due to their biodegradability, so also the synthetic ones which are synthesized from poly(lactide) (PLA) and its copolymers with glycolide, poly(lactic-co-glycolic acid) (PLGA) to give rise to poly (lactic acid) (PLA), poly (cyanoacrylates) (PACA), poly (acrylic acid), poly (anhydrides), poly(amides), poly (ortho esters), poly(ethylene glycol), and poly(vinyl alcohol) (PVA) , poly (isobutylnanoacrylate) (PIBCA), poly (ethylene oxide) (PEO), poly(α - caprolactone) (PCL) (Muhamad and Selvakumaran 2014; Álvarez-Paino *et al.*, 2017). Polymeric nanoparticles have been used as carrier molecules to deliver antimicrobials with promising therapeutic results (Zhang *et al.*, 2010).

Solid Lipid Nanoparticles (SLN)

These are nanoparticles which comprises of lipids that are solids at room temperature with a surfactant for emulsification. These lipids include fatty acids like triglycerides, steroids, partial glycerides and waxes. Surfactants that are commonly used as emulsifiers to stabilize lipid dispersion includes soybean lecithin, phosphatidylcholine, poloxamer 188, sodium cholate, and sodium glycocholate (Zhang *et al.*, 2010). They are more stable with longer release time when compared to liposomes and are safer than polymeric liposomes (Naseri *et al.*, 2015). SLN retains moisture in skin by preventing evaporation, this makes it useful in delivering pharmaceutical preparations meant for the skin due to its occlusive property (Zhang *et al.*, 2010). Although large scale production of SLN is cheap, it has a major disadvantage of low loading rates because of the crystalline nature of the SLN (Naseri *et al.*, 2015).

Dendrimer

These are “tree-like”, highly branched cascade molecules, comprising of a central stem from which branches arise from due to chemical reactions. Recently, this nanomolecule has been used in drug delivery, and the drug is loaded into the dendrimer either by encapsulation, complexation, or conjugation at the end of each branch (Abbasi *et al.*, 2014) like other nanoparticles, they have an advantage of increased

solubility and stability with low cytotoxicity. Increased addition of PEG to Poly (amido amine) PAMAM dendrimers decreases their toxicity (Lopez *et al.*, 2009)

Liposomes

Liposomes are small size spherically shaped vesicles made up of phospholipids, cholesterol and nontoxic surfactants which are used as carrier systems to deliver both hydrophilic and hydrophobic drug (Alavi *et al.*, 2017). Bangham in 1961 described liposomes as spherical vesicles quickly formed as a result of contact of phospholipids with water due to the bilayer nature of the lipids (Bangham *et al.*, 1965). "Liposome" is derived from two Greek word "lipo" and "soma" meaning fat and body, respectively (Anwekar *et al.*, 2011).

The spherical vesicles can be used as carriers to deliver drugs to target sites and these drug particles can either be encapsulated within the centre of the liposome (aqueous phase) or matrixed into the phospholipid bilayer. Advantages of liposomes includes carriers for both hydrophobic and hydrophilic drugs in one liposome, stability and prevents the decomposition of the drug. It reduces the toxicity of the drug and prevents exposure of sensitive tissue to toxic drugs and can be designed to release the drugs only at specific target site (Anwekar *et al.*, 2011 ; Akbarzadeh *et al.*, 2013). Despite this advantage, liposomes have short half-life, high production cost, leakage and fusion of encapsulated drug molecules and the covering phospholipid may undergo oxidation. Study of liposomes allows for improvement in drug delivery system and demonstrates its use in infectious diseases (Beaulac *et al.*, 1998; Drulis-Kawa *et al.*, 2009; Jung *et al.*, 2015) and cancer (Malam *et al.*, 2009) .

Structural contents of liposome

The two major structural components of liposomes are phospholipids and cholesterol. Glycerol containing phospholipids are the most commonly used phospholipids for liposome formulation. They represent greater than 50% of weight of lipids in biological membranes (Anwekar *et al.*, 2011). These phospholipids include Phosphatidyl choline (Lecithin) (PC), Phosphatidyl ethanolamine (cephalin) (PE), Phosphatidyl serine (PS), Phosphatidyl inositol (PI) and Phosphatidyl Glycerol (PG)Phosphatidic acid. Cholesterol is incorporated in the phospholipid in a high ratio of 1:1 or 2:1 of cholesterol to phosphatidyl choline with its hydroxyl group placed towards the aqueous phase and aliphatic chain placed parallel to the acyl chains in the centre of the phospholipid (Anwekar *et al.*, 2011).

Classification of liposomes

Liposomes can be classified structurally using their size and number of lipid bilayers. (Akbarzadeh *et al.*, 2013; Rani, 2013). They are classified as (1) Small unilamellar vesicles (SUV) (20 -100nm), (2)Medium sized unilamellar vesicles, (3) Unilamellar vesicles

(UV) (this could be in all size ranges), (4) Large unilamellar vesicles of (>100nm) (LUV), (5) Multilamellar large vesicles (MLV) (> 0.005 μ m), (6) Giant unilamellar vesicles (GUV) (> 1 μ m), (7) Oligolamellar lamellar Vesicles (OLV) and (8) Multivesicular vesicles (MVV) (Rani, 2013).

By composition and application, liposomes are classified as

Conventional/Ordinary liposomes which are composed of negatively or neutral charged phospholipids and cholesterol only that can be easily phagocytosed by the reticulo endothelial system (RES) (Paliwal *et al.*, 2015). The easy and fast removal of these liposomes from circulation by RES makes it possible to produce liposomes against parasites of the liver and spleen. Though the fast and early removal of the liposomes from circulation is a draw back as it makes them unable to be used to treat diseases for a long time (Storm and Crommelin, 1998).

P^H sensitive liposomes: These are liposomes made up of P^H sensitive lipid contents like cholesterol hemisuccinate (CHEMS), phosphatidyl ethanolamine (PE), oleic acid (OA) or dioleoylphosphatidyl ethanolamine (DOPE) (Karanth and Murthy, 2007). These vesicles fuse with cells and empty their contents into cytoplasm at low P^H , but are quickly recognized and removed by phagocytes of the RES. They addition of a hydrophilic polymer polyethylene glycol (PEG) increases their half-life and are of use in delivery of drugs in disease associated with low P^H conditions (Paliwal *et al.*, 2015).

Stealth (Stearically stabilized) liposomes: They are produced to make up for the disadvantage of quick elimination of conventional liposomes from the circulation by attaching polyethylene glycol (PEG) covalently to the outer surface of the liposome. The hydrated components of the PEG provide a barrier between the liposome and cellular environment (Storm and Crommelin, 1998; Karanth and Murthy, 2007).

Immunoliposomes are liposomes with antibodies or antibody fragments which facilitate binding at target site to enable them to bypass the adenosine triphosphate binding cassette transport so as to be internalized by endocytosis where the drug can be deposited near the nucleus (Tomuleasa *et al.*, 2014). This is important in overcoming resistance in cancer chemotherapy. PEG is usually added to it to prolong the lifespan of liposomes (Rani, 2013).

Cationic liposomes are made up of a positively charged lipid and a co or helper lipid like dioleoyl phosphatidyl ethanolamine (DOPE) and dioleoyl phosphatidylcholine (DOPC) (Rani, 2013). Interaction of the cationic lipid with DNA results in the formation of a cationic lipid DNA complex which provide protection and promote cellular internalization and expression of the condensed plasmid (Storm and Crommelin, 1998).

Fusogenic liposomes are liposomes made up of ultraviolet-inactivated Sendai virus and conventional liposomes. Fusogenic liposome targets the fusion mechanism of entry of the virus into the cell to deliver its content directly rather than depend on the been taken up by phagocytes of the RES as seen in the other liposomes. This has the advantage that it can cross with drugs into the cytoplasm or nucleus with minimal lysosomal enzyme interference. It is mostly used in the delivery of anticancer agents (Akbarzadeh *et al.*, 2013)

Based on the method of preparation, liposomes can be classified as (1) single or oligolamellar vesicles made by reverse phase evaporation method (REV), (2) multilamellar large vesicles made by reverse phase evaporation method (MLV-REV), (3) vesicles produced from the dehydration rehydration method (DRV), (4) stable plurilamellar vesicles (SPLV) (5) frozen and thawed multilamellar vesicles (FAT-MLV) (6) vesicles prepared by extrusion techniques (VET) (7) vesicles prepared by fusion (FUV), (8) vesicles prepared by French pressure cell (extrusion) (FPV) (Akbarzadeh *et al.*, 2013; Rani, 2013).

Calcium carbonate nanoparticles

Cockle shell is the waste by product derived from processing cockles or *anadara granosa*, a type of bivalve shellfish enjoyed as a delicacy in South East Asia (Mohamed *et al.*, 2012). The main component of the cockle shell is calcium carbonate (Bharatham *et al.*, 2014), an abundant biomaterial produced by a living organism which through biomedical engineering has been used as a source of biominerals for the treatment of bone lesions (Kiranda *et al.*, 2018). Synthesis of calcium carbonate nanoparticles (Nurul Islam *et al.*, 2013; Pan *et al.*, 2018) has shown promising results in the delivery of antibiotics (Kamba *et al.*, 2013; Isa *et al.*, 2016). There are four different forms, namely amorphous calcium carbonate (ACC), vaterite, aragonite and calcite (Bharatham *et al.*, 2014), with vaterite been the least thermodynamically stable and most soluble (Ni and Ratner, 2008; Weiss *et al.*, 2014). All the polymorphs are found in cockle shell with aragonite been the least abundant (Kamba *et al.*, 2013). During synthesis, all the 4 forms are affected by variables of P^H and temperature with P^H been the most significant (Weiss *et al.*, 2014). ACC phase is unstable and short-lived and serves as a precursor of crystal growth of the other polymorphs (Weiss *et al.*, 2014) Vaterite, also known as μ -CaCO₃ is the least common compared to aragonite and calcite because of its low thermodynamic stability. It can be converted to calcite and aragonite in aqueous solution (Ni and Ratner, 2008). Calcite is the most stable form of the four, it has a rhombohedral unit cell, found to form large single crystal cubic-like crystalline particles (Boyjoo *et al.*, 2014). Aragonite is thermodynamically unstable and an important biomedical tool, due to its denser nature when compared to calcite. Among the four, aragonite has been deeply researched into because of its high mechanical strength, biocompatibility and

biodegradability. Aragonite can be synthesized and its surface characteristics modified to ensure efficient drug loading and delivery (Kamba *et al.*, 2013; Jain *et al.*, 2014; Declat *et al.*, 2016; Isa *et al.*, 2016; Saidykhani *et al.*, 2016).

Summary of antimicrobial effect of nanoparticles

Gold nanoparticles have been used for treatment of experimental infections caused by *Corynebacterium pseudotuberculosis*, *Bacillus calmette Guerin*, *Escherichia coli*, *Klebsiella pneumonia*, *Staphylococcus aureus*, *Bacillus subtilis*, *S. typhimurium DT104*, and *S. aureus* in recent years (Zhou *et al.*, 2012; Mohamed *et al.*, 2017; Shamaila *et al.*, 2016; Shareena Dasari *et al.*, 2015; Payne *et al.*, 2016). Results from the research showed excellent antibacterial effect of gold against these microorganisms. Similarly, gold nanoparticles capped with Kanamycin enhanced the antibacterial effect of kanamycin as reported by Payne *et al.*, 2016). Loading antibiotics into silver nanoparticles resulted in synergistic effect of silver nanoparticles with antibiotics. None of the silver nanoparticle-antibiotic combinations demonstrated antagonistic effect. (Smekalova *et al.*, 2016). Commercially available zinc and titanium dioxide nanoparticles tested against biofilm producing methicillin-resistant staphylococcus aureus showed considerable antibacterial and antibiofilm activity (Jesline *et al.*, 2015). Testing *Escherichia coli*, *Staphylococcus aureus* and *Enterococcus faecalis* with copper nanoparticles loaded with amino acid chelates showed a significant enhanced antimicrobial activity of the copper chelates (Dealba-Montero *et al.*, 2017). PLGA-PLH-PEG nanoparticles demonstrated an increase in minimum inhibitory concentration against microorganisms when compared with PLGA-PEG nanoparticles loaded with vancomycin or vancomycin alone (Aleksandar *et al.*, 2012). Simeonova and Ivanova, (2017), reported that the antibacterial activity of ciprofloxacin- poly (butyl cyanoacrylate) nanoparticle combination was similar to that of ciprofloxacin alone. Nanoparticles increase the antimicrobial activity of antibiotics encapsulated in them and also reduces the amount of antibiotics required to inhibit micro organisms (Pavan Kumar Reddy *et al.*, 2014; Nicolosi *et al.*, 2015; Yegin *et al.*, 2016; Shadzly, 2017; Xie *et al.*, 2017; Severino *et al.*, 2017). The synergistic activity of nanoparticles loaded with antibiotics is enhanced because some nanoparticles possess antibacterial activity (Pignatello *et al.*, 2011 Aboutaleb *et al.*, 2012 ; Winnicka *et al.*, 2013; Vembu *et al.*, 2015; Cheung Lam *et al.*, 2016; Bartomeu *et al.*, 2017; Gholami *et al.*, 2017)

CONCLUSION

The unique characteristics of nanoparticles can be exploited in the treatment of bacterial infections where resistance have been developed against antibiotic drugs. The induction of oxidative stress in antibiotic resistant microorganisms which leads to its death has opened a new pathway in the treatment of bacterial

infections by nanoparticles whose causative agents have defied antibiotic treatment. The major problem with treatment in CLA is because antibiotics do not reach the organisms due to development of islets of infection walled away in pus, biofilm formation and the ability of *Corynebacterium pseudotuberculosis* to acquire resistance genes. Development of nanoparticles loaded with antibiotics may bring an end to the menace of antimicrobial resistance in the treatment of CLA therefore reducing the economic burden placed on livestock owners.

REFERENCES

- Abbasi, Elham, Sedigheh Fekri Aval, Abolfazl Akbarzadeh, Morteza Milani, and Hamid Tayefi Nasrabadi. 2014. "Dendrimers : Synthesis , Applications , and Properties" 9 (1): 1–10. doi:10.1186/1556-276X-9-247.
- Abdinasir Yusuf Osman, Faez Firdaus Jesse Abdullah, Eric Lim Teik Chung, Yusuf, Abdul Aziz Saharee Abba, Muhammad Abubakar Sadiq, Konto Mohammed, Mohd Azmi Mohd Lila and Abdul Wahid Haron,. 2015. "Caseous Lymphadenitis in a Goat: A Case Report,". doi:10.5455/ijlr.20150221071744.
- Aboutaleb, Ehsan, Massoumeh Noori, Narges Gandomi, Fatemeh Atyabi, and Mohammad Reza Fazeli. 2012. "Improved Antimycobacterial Activity of Rifampin Using Solid Lipid Nanoparticles," 1–8.
- Aderibigbe, Blessing Atim. 2017. "Metal-Based Nanoparticles for the Treatment of Infectious Diseases." doi:10.3390/molecules22081370.
- Aggarwal, Divya, and Ujjwal Nautiyal. 2016. "Ethosomes : A Review." *International Journal of Pharmaceutical and Medicinal Research* 4 (1): 354–63.
- Akbarzadeh, Abolfazl, Rogaie Rezaei-sadabady, Soodabeh Davaran, Sang Woo Joo, and Nosratollah Zarghami. 2013. "Liposome : Classification , Preparation , and Applications." *Nanoscale Research Letters* 8 (1). Nanoscale Research Letters: 1. doi:10.1186/1556-276X-8-102.
- Al-Gaabary, Magdy H., Salama A. Osman, and Atef F. Oreiby. 2009. "Caseous Lymphadenitis in Sheep and Goats: Clinical, Epidemiological and Preventive Studies." *Small Ruminant Research* 87 (1–3): 116–21. doi:10.1016/j.smallrumres.2009.10.008.
- Alavi, Mehran, Naser Karimi, and Mohsen Safaei. 2017. "Application of Various Types of Liposomes in Drug Delivery Systems." *Advanced Pharmaceutical Bulletin* 7 (1): 3–9. doi:10.15171/apb.2017.002.
- Aleksandar, F, Timothy K Lu, A Vlad, Christopher J Yoon, Robert Langer, C Omid, and Citable Link. 2012. "NIH Public Access." *aCS Nano* 6 (5): 4279–87. doi:10.1021/nn3008383.Surface.
- Analette I. Lopez, Rose Y. Reins, Alison M. McDermott, Barbara W. Trautner, And, and Chengzhi Cai. 2009. "Antibacterial Activity and Cytotoxicity of PEGylated Poly (Amidoamine) Dendrimers† Analette." *Molecular Biosystems* 5 (10): 1148–56. doi:10.1039/b904746h.
- Anwekar, Himanshu, Sitasharan Patel, and A K Singhai. 2011. "Liposome-as Drug Carriers." *Int. J. of Pharm. & Life Sci. (IJPLS)* 2 (7): 945–51. <http://www.ijplsjournal.com/issues/PDFfiles/july2011/12.pdf>.
- Aruguete, Deborah M., Bojeong Kim, Michael F. Hochella, Yanjun Ma, Yingwen Cheng, Andy Hoegh, Jie Liu, and Amy Pruden. 2013. "Antimicrobial Nanotechnology: Its Potential for the Effective Management of Microbial Drug Resistance and Implications for Research Needs in Microbial Nanotoxicology." *Environ. Sci.: Processes Impacts* 15 (1): 93–102. doi:10.1039/C2EM30692A.
- Bahadar, Haji, Faheem Maqbool, Kamal Niaz, and Mohammad Abdollahi. 2016. "Toxicity of Nanoparticles and an Overview of Current Experimental Models." *Iranian Biomedical Journal* 20 (1): 1–11. doi:10.7508/ibj.2016.01.001.
- Bangham, A D, M M Standish, and J C Watkins. 1965. "Diffusion of Univalent Ions across the Lamellae of Swollen Phospholipids." *Journal of Molecular Biology* 13 (1): 238–52. <http://www.ncbi.nlm.nih.gov/pubmed/5859039>.
- Bastos, Bruno Lopes, Ricardo Wagner Dias Portela, Fernanda Alves Dorella, Dayana Ribeiro, Nbia Seyffert, Thiago Luiz de Paula Castro, Anderson Miyoshi, Srgio

- Costa Oliveira, Roberto Meyer, and Vasco Azevedo. 2012. "Corynebacterium Pseudotuberculosis: Immunological Responses in Animal Models and Zoonotic Potential." *Journal of Clinical & Cellular Immunology* 1 (S4): 1–15. doi:10.4172/2155-9899.S4-005.
- Bharatham, Hemabarathy, Md Zuki Abu Bakar Zakaria, Enoch Kumar Perimal, Loqman Mohamad Yusof, and Muhajir Hamid. 2014. "Mineral and Physicochemical Evaluation of Cockle Shell (Anadara Granosa) and Other Selected Molluscan Shell as Potential Biomaterials." *Sains Malaysiana* 43 (7): 1023–29.
- Boyjoo, Yash, Vishnu K. Pareek, and Jian Liu. 2014. "Synthesis of Micro and Nano-Sized Calcium Carbonate Particles and Their Applications." *J. Mater. Chem. A* 2 (35). Royal Society of Chemistry: 14270–88. doi:10.1039/C4TA02070G.
- Cheung Lam, Annie H., Natalie Sandoval, Ritambhara Wadhwa, Janine Gilkes, Thai Q. Do, William Ernst, Su Ming Chiang, et al. 2016. "Assessment of Free Fatty Acids and Cholesteryl Esters Delivered in Liposomes as Novel Class of Antibiotic." *BMC Research Notes* 9 (1). BioMed Central: 1–11. doi:10.1186/s13104-016-2138-8.
- Dakal, Tikam Chand, Anu Kumar, Rita S. Majumdar, and Vinod Yadav. 2016. "Mechanistic Basis of Antimicrobial Actions of Silver Nanoparticles." *Frontiers in Microbiology* 7 (NOV): 1–17. doi:10.3389/fmicb.2016.01831.
- Danmaigoro, Abubakar, Gayathri Thevi Selvarajah, Mohd Hezmee Mohd Noor, Rozi Mahmud, and Md Zuki Abu Bakar Zakaria. 2017. "Development of Cockleshell (Anadara Granosa) Derived CaCO₃ Nanoparticle for Doxorubicin Delivery." *Journal of Computational and Theoretical Nanoscience* 14 (10): 5074–86. doi:10.1166/jctn.2017.6920.
- Dayem, Ahmed Abdal, Mohammed Kawser Hossain, Soo Bin Lee, Kyeongseok Kim, Subbroto Kumar Saha, Gwang-mo Yang, Hye Yeon Choi, and Ssang-goo Cho. 2017. "The Role of Reactive Oxygen Species (ROS) in the Biological Activities of Metallic Nanoparticles," 1–21. doi:10.3390/ijms18010120.
- Dealba-Montero, I, Jesús Guajardo-Pacheco, Elpidio Morales-Sánchez, Rene Araujo-Martínez, G M Loreda-Becerra, Gabriel-Alejandro Martínez-Castañón, Facundo Ruiz, and M E Compeán Jasso. 2017. "Antimicrobial Properties of Copper Nanoparticles and Amino Acid Chelated Copper Nanoparticles Produced by Using a Soya Extract." *Bioinorganic Chemistry and Applications*, Article ID 1064918. doi:10.1155/2017/1064918.
- Declet, A., E. Reyes, and O. M. Suárez. 2016. "Calcium Carbonate Precipitation: A Review of the Carbonate Crystallization Process and Applications in Bioinspired Composites." *Reviews on Advanced Materials Science* 44 (1): 87–107.
- Drulis-Kawa, Zuzanna, Agata Dorotkiewicz-Jach, Jerzy Gubernator, Grzegorz Gula, Tomasz Bocser, and Włodzimierz Doroszkiewicz. 2009. "The Interaction between Pseudomonas Aeruginosa Cells and Cationic PC:Chol:DOTAP Liposomal Vesicles versus Outer-Membrane Structure and Envelope Properties of Bacterial Cell." *International Journal of Pharmaceutics* 367 (1–2): 211–19. doi:10.1016/j.ijpharm.2008.09.043.
- Duangjit, Sureewan, Praneet Opanasopit, Theerasak Rojanarata, and Tanasait Ngawhirunpat. 2013. "Evaluation of Meloxicam-Loaded Cationic Transfersomes as Transdermal Drug Delivery Carriers" 14 (1). doi:10.1208/s12249-012-9904-2.
- Fard, Javad Khalili, Samira Jafari, and Mohammad Ali Eghbal. 2015. "A Review of Molecular Mechanisms Involved in Toxicity of Nanoparticles." *Advanced Pharmaceutical Bulletin* 5 (4): 447–54. doi:10.15171/apb.2015.061.
- Figueiredo, Alessandra, De Castro Nassar, Gabriela Terezinha Daniel, Regina Ruiz, Simone Miyashiro, Eloísa Maria Scannapieco, Juraci De Souza Neto, and Lilian Gregory. 2015. "Diagnostic Comparison of Corynebacterium Pseudotuberculosis through Microbiological Culture and PCR in Sheep Samples," 1–6. doi:10.1590/1808-1657000692013.
- Galvão, Cleber Eduardo, Stenio Perdigão Fragoso, Carina Elisei de Oliveira, Odinéia Forner, Renata Ribeiro Bastos Pereira, Cleber Oliveira Soares, and

- Grácia Maria Soares Rosinha. 2017. "Identification of New Corynebacterium Pseudotuberculosis Antigens by Immunoscreening of Gene Expression Library." *BMC Microbiology* 17 (1). BMC Microbiology: 202. doi:10.1186/s12866-017-1110-7.
- Gholami, Mitra, Rashin Mohammadi, Mohsen Arzanlou, Fakhraddin Akbari Dourbash, Ebrahim Kouhsari, Gharib Majidi, Seyed Mohsen Mohseni, and Shahram Nazari. 2017. "In Vitro Antibacterial Activity of Poly (Amidoamine) -G7 Dendrimer." *BMC Infectious Diseases*, 1–11. doi:10.1186/s12879-017-2513-7.
- Habuš, Josipa, Krešimir Matanović, Zrinka Štritof Majetić, Tomislav Rukavina, Ante Ćorić, Zoran Milas, Vilim Starešina, Branka Šeol Martinec, and Nenad Turk. 2015. "Comparison of the Epizootiological and Clinical Features of Caseous Lymphadenitis and Morel ' S Disease in Goats" 85 (2): 163–73.
- Hariharan, H, K P Tiwari, S Kumthekar, and D Thomas. 2015. "Serological Detection of Caseous Lymphadenitis in Sheep and Goats Using a Commercial ELISA in Grenada , West Indies" 2015. doi:10.5171/2015.473459.
- Hasan, Saba. 2015. "A Review on Nanoparticles : Their Synthesis and Types A Review on Nanoparticles : Their Synthesis and Types," no. March: 7–10.
- Ieda Maria Sapateiro Torres, Etienne Barbosa Bento, Larissa da Cunha Almeida, Luisa Zaiden Carvalho Martins de Sá, Eliana Martins Lima. 2012. "Preparation, Characterization and in Vitro Antimicrobial Activity of Liposomal Ceftazidime and Cefepime against." *Brazilian Journal of Microbiology*, 984–92.
- Isa, Tijani, Zuki Abu Bakar Zakaria, Yaya Rukayadi, Mohd Noor Mohd Hezme, Alhaji Zubair Jaji, Mustapha Umar Imam, Nahidah Ibrahim Hammad, and Saffanah Khuder Mahmood. 2016. "Antibacterial Activity of Ciprofloxacin-Encapsulated Cockle Shells Calcium Carbonate (Aragonite) Nanoparticles and Its Biocompatibility in Macrophage J774A.1." *International Journal of Molecular Sciences* 17 (5). Multidisciplinary Digital Publishing Institute (MDPI). doi:10.3390/ijms17050713.
- Jain, Keerti, Neelesh Kumar Mehra, and Narendra Kumar Jain. 2014. "Potentials and Emerging Trends in Nanopharmacology." *Current Opinion in Pharmacology* 15 (1). Elsevier Ltd: 97–106. doi:10.1016/j.coph.2014.01.006.
- Jesline, A, Neetu, P John, P M Narayanan, C Vani, and Sevanan Murugan. 2015. "Antimicrobial Activity of Zinc and Titanium Dioxide Nanoparticles against Biofilm-Producing Methicillin-Resistant Staphylococcus Aureus." doi:10.1007/s13204-014-0301-x.
- Jesse Faez Firdaus Abdullah, Yusuf Abba, S. R. nurul, Muhammad Abubakar Sadiq , lawan Adamu, Asinamai Athliamai Bitrus, Eric Lim Teik Chung, Mohammed Azmi Idris Umar Hambali, Wahid Haron, and Mohammed Lila. 2017. "clinical case of caseous lymphadenitis in a goat: case management." *Malaysian Journal of Veterinary Research*, 31–35.
- Jones, David E., Hamidreza Ghandehari, and Julio C. Facelli. 2015. "Predicting Cytotoxicity of PAMAM Dendrimers Using Molecular Descriptors." *Beilstein Journal of Nanotechnology* 6 (1): 1886–96. doi:10.3762/bjnano.6.192.
- Jung, Sung Woo, Soracha Thamphiwatana, Liangfang Zhang, and Marygorret Obonyo. 2015. "Mechanism of Antibacterial Activity of Liposomal Linolenic Acid against Helicobacter Pylori," 1–13. doi:10.1371/journal.pone.0116519.
- Kamba, Shafiu, Maznah Ismail, Samer Hussein-Al-Ali, Tengku Ibrahim, and Zuki Zakaria. 2013. "In Vitro Delivery and Controlled Release of Doxorubicin for Targeting Osteosarcoma Bone Cancer." *Molecules* 18 (9): 10580–98. doi:10.3390/molecules180910580.
- Karanth, H., and R. S. R. Murthy. 2007. "pH-Sensitive Liposomes-Principle and Application in Cancer Therapy." *Journal of Pharmacy and Pharmacology* 59 (4): 469–83. doi:10.1211/jpp.59.4.0001.
- Khan, Mohd Farhan, Akhter H Ansari, M Hameedullah, Ejaz Ahmad, Mohammad Mezbaul Alam, Abu Mustafa Khan, Zeid A Alothman, and Iqbal Ahmad. 2016. "Sol-Gel Synthesis of Thorn-like ZnO Nanoparticles Endorsing Mechanical

- Stirring Effect and Their Antimicrobial Activities: Potential Role as Nano-Antibiotics.” *Science Reports* 6. Nature Publishing Group: 1–12. doi:10.1038/srep27689.
- Khan, Nida Tabassum. 2017. “Nanoparticles Mediated Drug Delivery.” *Journal of Pharmacogenomics & Pharmacoproteomics* 8 (3): 8–10. doi:10.4172/2153-0645.1000172.
- Khanna, Puja, Cynthia Ong, Boon Huat Bay, and Gyeong Hun Baeg. 2015. “Nanotoxicity: An Interplay of Oxidative Stress, Inflammation and Cell Death,” 1163–80. doi:10.3390/nano5031163.
- Kiranda, H. K., Mahmud, R. Danmaigoro, A. and Zuki A. Z. (2018). Fabrication, Characterization and Cytotoxicity of Spherical-Shaped Conjugated Gold-Cockle Shell Derived Calcium Carbonate Nanoparticles for Biomedical Applications. *Nanoscale Research Letters*, 13:1-10
- Mahmood, Z.K.H., 1F.F. Jesse, 1A.A. Saharee, 2S. Jasni, 1R. Yusoff and 1H. Wahid 1Department. 2017. “Clinio-Pathological Changes in Goats Challenged with *Corynebacterium Pseudotuberculosis* and Its Exotoxin (PLD).” Accessed November 20. <http://thescipub.com/PDF/ajavsp.2015.12.132.pdf>.
- Malam, Yogeshkumar, Marilena Loizidou, and Alexander M. Seifalian. 2009. “Liposomes and Nanoparticles: Nanosized Vehicles for Drug Delivery in Cancer.” *Trends in Pharmacological Sciences* 30 (11): 592–99. doi:10.1016/j.tips.2009.08.004.
- Mallakpour, S, and V Behranvand. 2016. “Polymeric Nanoparticles: Recent Development in Synthesis and Application” 10 (11): 895–913. doi:10.3144/expresspolymlett.2016.84.
- Marta, Á, Alexandra Muñoz-bonilla, and Marta Fern. 2017. “Antimicrobial Polymers in the Nano-World” 7 (2): 1–44. doi:10.3390/nano7020048.
- Maynard, Andrew D., David B. Warheit, and Martin A. Philbert. 2011. “The New Toxicology of Sophisticated Materials: Nanotoxicology and beyond.” *Toxicological Sciences* 120 (SUPPL.1). doi:10.1093/toxsci/kfq372.
- Mohamed, Marwah M., Shereen A. Fouad, Hisham A. Elshoky, Gina M. Mohammed, and Taher A. Salaheldin. 2017. “Antibacterial Effect of Gold Nanoparticles against *Corynebacterium Pseudotuberculosis*.” *International Journal of Veterinary Science and Medicine* 5 (1). Elsevier: 23–29. doi:10.1016/j.ijvsm.2017.02.003.
- Mohd Abd Ghafar, Syairah Liyana, Mohd Zobir Hussein, and Zuki Abu Bakar Zakaria. 2017. “Synthesis and Characterization of Cockle Shell-Based Calcium Carbonate Aragonite Polymorph Nanoparticles with Surface Functionalization.” *Journal of Nanoparticles* 2017 (January). Hindawi: 1–12. doi:10.1155/2017/8196172.
- Muhamad, Ida Idayu, and Suguna Selvakumaran. 2014. “Designing Polymeric Nanoparticles for Targeted Drug Delivery System Outline :” *Nanomedicine* 11: 287–313.
- Mustakimah Mohamed, Suzan yusup, and Saikat Maitra. 2012. “Decomposition Study of Calcium Carbonate in Cockle Shell.” *Journal of Engineering Sciencee and Technology* 711: 1–10.
- Muthusamy, K, and N A Sabri. 2017. “Cockle shell: A Potential Partial Coarse Aggregate Replacement in Concrete.” Accessed December 12. <http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.301.5805&rep=rep1&type=pdf>.
- Ni, Ming, and Buddy D Ratner. 2008. “Differentiation of Calcium Carbonate Polymorphs by Surface Analysis Technique - An XPS and TOF-SIMS Study.” *National Institute of Health* 40 (10): 1356–61. doi:10.1002/sia.2904.Differentiation.
- Nicolosi, Daria, Sarha Cupri, Carlo Genovese, Gianna Tempera, Roberto Mattina, and Rosario Pignatello. 2015. “Nanotechnology Approaches for Antibacterial Drug Delivery: Preparation and Microbiological Evaluation of Fusogenic Liposomes Carrying Fusidic Acid.” *International Journal of Antimicrobial Agents* 45 (6). Elsevier: 622–26. doi:10.1016/j.ijantimicag.2015.01.016.

- Nurul Islam, Kh, Md Equb Ali, Md Zuki Bin Abu Bakar, MY Loqman, Aminul Islam, Md Saiful Islam, Md Mahfujur Rahman, and Mahbub Ullah. 2013. "A Novel Catalytic Method for the Synthesis of Spherical Aragonite Nanoparticles from Cockle Shells." *Powder Technology* 246. doi:10.1016/j.powtec.2013.05.046.
- Olender, Alina. 2012. "Mechanisms of Antibiotic Resistance in Corynebacterium Spp. Causing Infections in People." *Antibiotic Resistant Bacteria – A Continuous Challenge in the New Millennium*, 387–402.
<http://www.intechopen.com/books/antibiotic-resistant-bacteria-a-continuous-challenge-in-the-new-millennium/mechanisms-of-antibiotic-resistance-in-corynebacterium-spp-causing-infections-in-people%5CnInTech>.
- Oreiby, Atef F. 2015. "Diagnosis of Caseous Lymphadenitis in Sheep and Goat." *Small Ruminant Research* 123 (1). Elsevier B.V.: 160–66. doi:10.1016/j.smallrumres.2014.11.013.
- Othman, M, F F Jesse, L Adamu, Y Abba, Adza Rina M N, A A Saharee, A H Wahid, and Zamri Saad M. 2014. "Changes in Serum Progesterone and Estrogen Concentrations in Non-Pregnant Boer Does Following Experimental Infection with Corynebacterium Pseudotuberculosis Changes in Serum Progesterone and Estrogen Concentrations in Non-Pregnant Boer Does Following Expe." *Journal of Veterinary Advances* 4 (5): 524–28.
- Paliwal, Shivani Rai, Rishi Paliwal, and Suresh P. Vyas. 2015. "A Review of Mechanistic Insight and Application of pH-Sensitive Liposomes in Drug Delivery." *Drug Delivery* 22 (3): 231–42. doi:10.3109/10717544.2014.882469.
- Palza, Humberto. 2015. "Antimicrobial Polymers with Metal Nanoparticles," 2099–2116. doi:10.3390/ijms16012099.
- Parashar, Tarun, Soniya, Roopesh Sachan, Vishal Singh, Gaurav Singh, Satyanand Tyagi, Chirag Patel, and Anil Gupta. 2013. "Review Article Ethosomes : A Recent Vesicle of Transdermal Drug Delivery System." *International Journal of Research and Development in Pharmacy and Life Sciences* 2 (2): 285–92.
- Pavan kumar reddy A, S. Parthiban, A. Vikneswari, G.P. Senthilkumar. 2014. "Evaluation of Potential Antimicrobial Activity of Levofloxacin." *Asian Journal of Research in Biological and Pharmaceutical Sciences* 2 (3): 99–111.
- Payne, Jason N., Hitesh K. Waghwani, Michael G. Connor, William Hamilton, Sarah Tockstein, Harsh Moolani, Fenil Chavda, Vivek Badwaik, Matthew B. Lawrenz, and Rajalingam Dakshinamurthy. 2016. "Novel Synthesis of Kanamycin Conjugated Gold Nanoparticles with Potent Antibacterial Activity." *Frontiers in Microbiology* 7 (MAY): 1–10. doi:10.3389/fmicb.2016.00607.
- Pignatello, Rosario, Daria Nicolosi, and Vito Mar Nicolosi. 2011. "Fusogenic Liposomes as New Carriers to Enlarge the Spectrum of Action of Antibiotic Drugs against Gram-Negative Bacteria." *Science against Microbial Pathogens: Communicating Current Research and Technological Advances* 1: 52–60.
- Pujalté, Igor, Isabelle Passagne, Brigitte Brouillaud, Mona Tréguer, Etienne Durand, Céline Ohayon-Courtès, and Béatrice L'Azou. 2014. "Cytotoxicity and Oxidative Stress Induced by Different Metallic Nanoparticles on Human Kidney Cells." *Beilstein Journal of Nanotechnology* 5 (1): 1590–1602. doi:10.1186/1743-8977-8-10.
- R. Ladj, ab A. Bitar, a M. Eissa,*ac Y. Mugnier, b R. Le Dantec, b H. Fessia and A. Elaissaria. 2013. "Journal of Materials Chemistry B." *Journal of Materials Chemistry B*, no. 1: 1381–96. doi:10.1039/c2tb00301e.
- Rani, Dash Tapaswi. 2013. "Liposome as a Potential Drug Delivery System : A Review." *International Research Journal of Pharmacy* 4 (1): 6–12.
- Ravi, Kumar, Manvir Singh, and A C Rana. 2012. "ISSN 2230 – 8407 Transferosomes : A Novel Approach For Transdermal Drug Delivery." *International Research Journal of Pharmacy* 3 (1): 20–24.
- Ribeiro, Dayana, Fernanda Alves Dorella, Luis Gustavo, Carvalho Pacheco, Núbia Seyffert, Thiago Luiz, De Paula Castro, et al. 2013. "Bacteriology & Parasitology Subclinical Diagnosis of Caseous Lymphadenitis Based on

- ELISA in Sheep from Brazil” 4. doi:10.4172/2155-9597.1000170.
- Saidykhan, Lamin, Md Zuki Bin Abu Bakar, Yaya Rukayadi, Aminu Umar Kura, and Saiful Yazan Latifah. 2016. “Development of Nanoantibiotic Delivery System Using Cockle Shell-Derived Aragonite Nanoparticles for Treatment of Osteomyelitis.” *International Journal of Nanomedicine* 11. Dove Press: 661–73. doi:10.2147/IJN.S95885.
- Sandhirakasu Vembu, Srinivasan Pazhamalai, Mannathusamy Gopalakrishnan et al. 2015. “Potential Antibacterial Activity of Triazine Dendrimer: Synthesis and Controllable Drug Release Properties.” *Biorganic and Medicinal Chemistry* 23: 4561–66.
- Severino, Patrícia, Elisânia F Silveira, Kahynna Loureiro, Marco V Chaud, Danilo Antonini, Marcelo Lancellotti, Victor Hugo, et al. 2017. “European Journal of Pharmaceutical Sciences Antimicrobial Activity of Polymyxin-Loaded Solid Lipid Nanoparticles (PLX- SLN): Characterization of Physicochemical Properties and in Vitro E Ffi Cacy.” *European Journal of Pharmaceutical Sciences* 106 (May). Elsevier: 177–84. doi:10.1016/j.ejps.2017.05.063.
- Shamaila, Shahzadi, Noshin Zafar, Saira Riaz, Rehana Sharif, Jawad Nazir, and Shahzad Naseem. 2016. “Gold Nanoparticles: An Efficient Antimicrobial Agent against Enteric Bacterial Human Pathogen.” *Nanomaterials* 6: 71–81. doi:10.3390/nano6040071.
- Shareena Dasari, T P, Y Zhang, and H Yu. 2015. “Antibacterial Activity and Cytotoxicity of Gold (I) and (III) Ions and Gold Nanoparticles.” *Biochemistry & Pharmacology* 4 (6): 1–16. doi:10.4172/2167-0501.1000199.
- Shazly, Gamal A. 2017. “Ciprofloxacin Controlled-Solid Lipid Nanoparticles : Characterization , In Vitro Release , and Antibacterial Activity Assessment” 2017.
- Simeonova, Margarita Y, and Galya Ivanova. 2017. “International Journal of Nanomaterials , Nanotechnology and Nanomedicine Synthesis , Characterization and Antibacterial Activity of Ciprofloxacin Loaded Polymer Nanoparticles for Parenteral Application” 3: 34–43.
- Smekalova, Monika, Virginia Aragon, Ales Panacek, Robert Pucek, Radek Zboril, and Libor Kvitek. 2016. “Enhanced Antibacterial Effect of Antibiotics in Combination with Silver Nanoparticles against Animal Pathogens” 209: 174–79. doi:10.1016/j.tvj.2015.10.032.
- Storm, Gert and Crommelin, J.A. 1998. “Liposome:quo Vadis?” *Research Focus Reviews* 1 (19–31). <https://www.chem.uwec.edu/Chem455/1ipo.pdf>.
- Syeda Saniya Fatima*1, Shireen Begum2, Talath Fatima1, Madiha Jabeen1, and IB. 2017. “Niosomes as Nanoparticulate Drug Carriers Syeda.” *International Journal of Pharmacy and Pharmaceutical Research* 9 (3): 117–33.
- Tomuleasa, Ciprian, Cornelia Braicu, Alexandra Irimie, Lucian Craciun, and Ioana Berindan-Neagoe. 2014. “Nanopharmacology in Translational Hematology and Oncology.” *International Journal of Nanomedicine* 9 (1): 3465–79. doi:10.2147/IJN.S60488.
- Wallace, David, and David R. 2015. “Nanotoxicology and Metalloestrogens: Possible Involvement in Breast Cancer.” *Toxics* 3 (4). Multidisciplinary Digital Publishing Institute: 390–413. doi:10.3390/toxics3040390.
- Wang, Linlin, Chen Hu, and Longquan Shao. 2017. “The Antimicrobial Activity of Nanoparticles: Present Situation and Prospects for the Future.” *International Journal of Nanomedicine* 12: 1227–49. doi:10.2147/IJN.S121956.
- Washburn, K E, V R Fajt, S D Lawhon, L G Adams, L A Tell, and W T Bissett. 2013. “Caprine Abscess Model of Tulathromycin Concentrations in Interstitial Fluid from Tissue Chambers Inoculated with *Corynebacterium Pseudotuberculosis* Following Subcutaneous or.” *Antimicrobial Agents and Chemotherapy* 57 (12): 6295–6304. doi:10.1128/AAC.00936-13.
- Washburn, Kevin E., Wesley T. Bissett, Virginia R. Fajt, Melissa C. Libal, Geoffrey T. Fosgate, Joseph A. Miga, and Kristine M. Rockey. 2009. “Comparison of Three

- Treatment Regimens for Sheep and Goats with Caseous Lymphadenitis.” *Journal of the American Veterinary Medical Association* 234 (9). American Veterinary Medical Association 1931 North Meacham Road - Suite 100, Schaumburg, IL 60173 USA 847-925-8070 847-925-1329 avmajournals@avma.org : 1162–66. doi:10.2460/javma.234.9.1162.
- Washburn, Kevin E.(2018). Caseous lymphadenitis in sheep and goats. The Merck manual online. Retrieved from The Merck Manuals Online Medical Library database.
- Weiss, Charles a, Kevin Torres-Cancel, Robert D Moser, P G Allison, E Rae Gore, Mei Q Chandler, and Philip G Malone. 2014. “Influence of Temperature on Calcium Carbonate Polymorph Formed from Ammonium Carbonate and Calcium Acetate.” *Journal of Nanotechnology and Smart Materials* 1: 1–6.
- Williamson, L H. 2001. “Caseous Lymphadenitis in Small Ruminants.” *The Veterinary Clinics of North America. Food Animal Practice* 17 (2): 359–71, vii. <http://www.ncbi.nlm.nih.gov/pubmed/11515406>.
- Winnicka, Katarzyna, Magdalena Wroblewska, Piotr Wieczorek, Pawel Tomasz Sacha, and Elzbieta Anna Tryniszewska. 2013. “The Effect of PAMAM Dendrimers on the Antibacterial Activity of Antibiotics with Different Water Solubility.” *Molecules* 18: 8607–17. doi:10.3390/molecules18078607.
- Xie, Shuyu, Fei Yang, Yanfei Tao, Dongmei Chen, Wei Qu, Lingli Huang, and Zhenli Liu. 2017. “Enhanced Intracellular Delivery and Antibacterial Efficacy of Enrofloxacin-Loaded Docosanoic Acid Solid Lipid Nanoparticles against Intracellular Salmonella.” *Nature Publishing Group*, no. August 2016. Nature Publishing Group: 1–9. doi:10.1038/srep41104.
- Yalçın Enis, İpek, Merve Küçükali Öztürk, Hande Sezgin, and Ezgi Ismar. 2017. “An Overview of Nanotoxicology.” *Engineering Sciences (NWSAENS)* 12 (1): 57–65. doi:10.12739/NWSA.2017.12.1.1A0373.
- Yasam, Venkata Ramesh, Satya Lavanya Jakki, Jawahar Natarajan, and Gowthamarajan Kuppusamy. 2014. “A Review on Novel Vesicular Drug Delivery: Proniosomes.” *Drug Delivery* 21 (4): 243–49. doi:10.3109/10717544.2013.841783.
- Yegin, Yagmur, Keila L Perez-lewis, Ming Zhang, Mustafa Akbulut, and Thomas M Taylor. 2016. “Development and Characterization of Geraniol-Loaded Polymeric Nanoparticles with Antimicrobial Activity against Foodborne Bacterial Pathogens.” *Journal of Food Engineering* 170. Elsevier Ltd: 64–71. doi:10.1016/j.jfoodeng.2015.09.017.
- Zhang, L, D Pornpattananangku, C-M J Hu, and C-M Huang. 2010. “Development of Nanoparticles for Antimicrobial Drug Delivery.” *Current Medicinal Chemistry* 17 (6): 585–94. <http://www.ncbi.nlm.nih.gov/pubmed/20015030>.
- Zhang, L, D Pornpattananangkul, C J Hu, and C Huang. 2010. “Development of Nanoparticles for Antimicrobial Drug Delivery.” *Current Medinal Chemistry* 17: 585–94.
- Zhou, Yan, Ying Kong, Subrata Kundu, Jeffrey D Cirillo, and Hong Liang. 2012. “Antibacterial Activities of Gold and Silver Nanoparticles against Escherichia Coli and Bacillus Calmette-Guérin.” *Journal of Nanobiotechnology* 10: 2–9. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3405418/pdf/1477-3155-10-19.pdf>.